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CORRESPONDENCE

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Purification of *M. leprae* Isolated from Human Skin Biopsies

TO THE EDITOR:

For many years after its discovery in 1873 (³), biopsies of nodules from lepromatous leprosy patients constituted the only source of the non-cultivable bacillus, Mycobacterium leprae. Draper's method (1) is currently being used by the WHO-IMMLEP program for the isolation of large quantities of M. leprae from armadillo tissues. We used the same protocol to isolate M. leprae from human skin biopsies. The data presented in Tables 1 and 2 show that Protocol 1/79 can be adapted for human tissue, although its reproducibility was not totally satisfactory. The often low bacterial counts in skin tissues and the relatively massive amounts of collagenous tissues and fats greatly affect the recovery, especially at the density gradient centrifugation step. However, the isolated M. leprae cells from skin tissues are free from the brownish contaminants which are readily found in isolates from armadillo liver tissues. Moreover, other enzymes and protein contaminants were not detected by starch gel electrophoresis, nor were they seen by light and electron microscopy.

The type of disease in the patient from whom the starting material is obtained affects the percentage recovery of *M. leprae* (Table 2). The *M. leprae* cells in the old LL cases could have been in the disintegrated form and could have been lost during the purification process. Skin biopsies from new LL cases and BL patients gave percentages of recovery comparable to those from armadillo liver tissues. Skin biopsies from patients with LL with reactivation and LL with ENL gave very good yields of pure *M. leprae* cells.

Preliminary observations suggest that M. leprae isolated from human skin biopsies may be more specific than those isolated

TABLE 1.	Yield of bacilli	and presence	e of tissue	contaminants	at different	stages of the
purification p	process.					

Durifornian	No. o	Presence of tissues				
Purification step	Expt. 1 ^a	Expt. 2 ^a	Expt. 3 ^a	Expt. 4 ^b	LM ^c	EM ^d
Skin tissue homo- genates	90 (100%)	650 (100%)	600 (100%)	66 (100%)	++++¢	++++
End of homogenization End of enzymatic	78 (86%)	480 (74%)	520 (87%)	47 (71%)	+++	++
treatment End of gradient	66 (73%)	420 (65%)	490 (82%)	39 (59%)	++	· +,
centrifugation End of two-phase	51 (57%)	390 (60%)	420 (70%)	27 (41%)	+	_f
separation	48 (53%)	280 (43%)	310 (52%)	21 (32%)	±	_

^a Expt. 1-3 = Human skin biopsies (pooled).

 d EM = Electron microscopy.

^b Expt. 4 = Mouse foot-pad tissue (pooled).
^c LM = Light microscopy.

• + = Presence of tissue contaminants.

f - = Absence of tissue contaminants.

475

International Journal of Leprosy

	Biopsy pool	Tissue - homogenates (×10 ⁵ AFB ml ⁻¹) (a)	Two-phase separation (×10 ⁵ AFB ml ⁻¹)		% Recovery
Disease type ^a	no. of specimens		Lower layer (b)	Upper layer (c)	$\left(\frac{a}{a} \times 100\%\right)$
LL old cases	5	8.9	0.0	0.0	_
LL new cases	8	281.0	0.0	131.1	47
LL with reactivation	4	131.4	3.1	87.6	67
LL with ENL	4	8.1	0.0	7.0	86
BL	7	71.0	0.0	31.4	44
TT	4	0.0	0.0	0.0	_
BT	3	0.0	0.0	0.0	-
Mouse foot pad	6	3.1	0.0	0.8	26

TABLE 2. Recovery of M. leprae isolated from skin biopsies taken from patients with different types of disease.

^a LL = lepromatous leprosy; ENL = erythema nodosum leprosum; BL = borderline lepromatous; TT = tuberculoid leprosy; BT = borderline tuberculoid.

from armadillo liver tissues (supplied by IMMLEP) in that the former did not react with any of the tuberculosis patients' sera tested, while the latter reacted with 5%-9% of these sera (²).

-Sen-Chiew Gan, M.Sc., Dip. Ed., Grad.D.B.A., C.Biol., M.I.Biol., A.I.B.A., A.M.S.A.

Research Officer, Tropical Diseases Institute for Medical Research 50588 Kuala Lumpur Malaysia

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1986

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