Determination of the D and L Configuration of Phenolic Glycolipids of *M. leprae* and *M. bovis*

TO THE EDITOR:

It has been reported by Hunter, et al. that the structure of the sugar part of the speciesspecific phenolic glycolipid-I (PGL-I) of Mycobacterium leprae is O- (3,6-di-Omethyl- β -D-glucopyranosyl)- $(1 \rightarrow 4)$ -O-(2,3)di-O-methyl- α -L-rhamnopyranosyl)-(1 \rightarrow 2)-3-O-methyl- α -L-rhamnopyranose (^{15, 16}). This structure has generally been accepted and used elsewhere. However, the absolute structure of each sugar has only been assumed and has not been determined in that report (16). Several laboratories have shown by synthetic study that the disaccharide and trisaccharide of PGL-I synthesized from Dglucose and L-rhamnose have almost the same activity as that of PGL-I (1-4, 6-10). This suggests that the assumption of Hunter, et al, is correct, but there has been no direct evidence concerning the absolute configurations of the sugar residues. On the other hand, Demarteau-Ginsberg and Lederer have reported that the structure of the sugar part of the PGL of M. bovis (mycoside B) is 2-O-methyl-D-rhamnose, which was determined by an optical rotation study (5). However, D-rhamnose is a very rare sugar, and the reports which appeared after that paper did not treat the absolute configuration of the sugar residue (11, 12). Therefore, it is necessary to make sure of this determination by a more direct method. This paper provides the gas chromatographic determination of the absolute structure of the sugar residues of the PGLs of M. leprae and M. bovis.

Analytical procedures of the absolute configuration were based on the glycosidation with optically active alcohol, (+)-2-butanol (¹³). Five hundred μ g of PGL-I from human-*M. leprae*-infected armadillo liver,

synthesized trisaccharide, p-(2-methoxycarbonylethyl)phenyl O-(3,6-di-O-methyl- β -D-glucopyranosyl)-(1 \rightarrow 4)-O-(2,3-di-Omethyl- α -L-rhamnopyranosyl)- $(1 \rightarrow 2)$ -3-Omethyl- α -L-rhamnopyranoside (9), or synthesized 2-O-methyl-L-rhamnopyranose was heated for 15 hr in 0.5 ml of (\pm) -2butanol or (+)-2-butanol in the presence of 25 mg of powdered Amberlite IR 120 (H⁺). The mixture was filtered, evaporated with the repeated addition of methanol, and then dissolved in 1 ml of methanol. Insoluble materials were filtered out with the aid of Celite. It was evaporated and dried. The residue was trimethylsilylated with 0.1 ml of TMS-PZ (Tokyo Kasei Co.) by heating the mixture at 40°C for 30 min, and an aliquot was analyzed by a capillary gas chromatograph (Hitachi G3000). Gas-liquid chromatography (GLC) conditions were as follows: Column: chemical bonded OV-1: d.f. $0.5 \,\mu\text{m}$; length 5 m (0.25 mm i.d.); temp. program = 150°C for 3 min then \rightarrow 200°C at 4°C/min; carrier gas = 34 cm/min; split ratio = 25:1; injection temp. = 220° C.

Figure 1 shows the results of GLC of the

FIG. 1. GLC of the sugar derivatives of the PGL-I of *M. leprae*: (+)-butanol-treated PGL-I (a), (\pm) -butanol-treated PGL-I (b), and (+)-butanol-treated synthetic trisaccharide (c) were trimethylsilylated and subjected to capillary GLC. (GLC conditions are given in the text.)

FIG. 2. GLC of the sugar derivatives of the phenolic glycolipid of *M. bovis*: (+)-butanol-treated PGL (a), (\pm)-butanol-treated PGL (b), and (+)-butanol-treated 2-*O*-methyl-L-rhamnopyranose (c) were trimethyl-silylated and subjected to capillary GLC.





sugar derivatives of the PGL-I of M. leprae. GLC of the sugar derivatives of PGL-I treated with (\pm) -2-butanol showed four 1:1 doublet-peaks corresponding to the derivatives of 2,3-di-O-methyl-D- and L-rhamnose (5.10 min and 5.30 min, (+)-2-butyl 2,3-di-O-methyl-D-rhamnoside should have the same retention time as (-)-2-butyl 2,3di-O-methyl-L-rhamnoside); 3-O-methyl-Dand L-rhamnose (6.10 and 6.25 min); 3,6di-O-methyl- α -D- and β -L-glucose (10.60 and 10.80 min); and 3,6-di-O-methyl-α-Land β -D-glucose (11.45 and 11.75 min). In the case of PGL-I treated with (+)-2-butanol, four singlet-peaks were observed at 5.30, 6.27, 10.60, and 11.80 min. Synthesized trisaccharide gave four singlet-peaks at 5.30, 6.25, 10.58, and 11.75 min and one additional singlet-peak at 8.95 min, which corresponded to p-hydroxyphenylpropionate. Cochromatography of the (+)-2-butanol-treated PGL and the (+)-2-butanoltreated synthesized trisaccharide gave the same pattern as that of (+)-2-butanol-treated PGL, except that the peak was at 8.95 min (p-hydroxyphenylpropionate). Therefore, the absolute configurations of the three sugars of M. leprae PGL-I were D for 3,6di-O-methylglucose and L for both 2,3-di-O-methylrhamnose and 3-O-methylrhamnose.

Figure 2 shows the results of GLC of the sugar derivatives of the PGL of M. bovis. GLC of the (\pm) -2-butanol-treated PGL shows only one doublet-peak at 7.60 and 7.80 min, which corresponds to the derivatives of 2,3-di-O-methyl-D- and L-rhamnose, respectively. In the case of the PGL treated with (+)-2-butanol, only one singlet-peak was observed at 7.80 min, which was completely in accord with that of (+)-2-butanol-treated 2-O-methyl-L-rhamnose. Cochromatography of (+)-2-butanol-treated PGL and (+)-2-butanol-treated 2-Omethyl-L-rhamnose gave the same pattern as that of (+)-2-butanol-treated PGL, showing that 2-O-methylrhamnose of M. bovis was in the L-configuration.

-Tsuyoshi Fujiwara, Ph.D.

Laboratory of Chemistry Institute for Natural Science Nara University Horai-cho 1230 Nara 631, Japan

REFERENCES

- BRENNAN, P. J. The phthiocerol-containing surface lipids of *Mycobacterium leprae*—a perspective of past and present work. Int. J. Lepr. **51** (1983) 387– 396.
- BRETT, S. J., DRAPER, P., PAYNE, S. N. and REES, R. J. W. Serological activity of a characteristic phenolic glycolipid from *Mycobacterium leprae* in sera from patients with leprosy and tuberculosis. Clin. Exp. Immunol. **52** (1983) 271–279.
- 3. CHATTERJEE, D., CHO, S.-N. and BRENNAN, P. J. Chemical synthesis and seroreactivity of O-(3,6di-O-methyl- β -D-glucopyranosyl)-(1 \rightarrow 4)-O-(2,3di-O-methyl- α -L-rhamnopyranosyl)-(1 \rightarrow 9)—oxynonanoyl-bovine serum albumin—the leprosyspecific, natural disaccharide-octyl-neoglycoprotein. Carbohydr. Res. **156** (1986) 39–56.
- CHO, S.-N., FUJIWARA, T., HUNTER, S. W., REA, T. H., GERBER, R. H. and BRENNAN, P. J. Use of artificial antigen containing the 3,6-di-*O*-methylβ-D-glucopyranosyl epitope for the serodiagnosis of leprosy. J. Infect. Dis. **150** (1984) 311–322.
- DEMARTEAU-GINSBERG, H. and LEDERER, E. Studies on the chemical structure of mycoside B. Biochem. Biophys. Acta 70 (1963) 442–451.
- FUJIWARA, T., ASPINALL, G. O., HUNTER, S. W. and BRENNAN, P. J. Chemical synthesis of the trisaccharide unit of the species-specific phenolic glycolipid from *Mycobacterium leprae*. Carbohydr. Res. 163 (1987) 41–52.
- FUJIWARA, T., HUNTER, S. W. and BRENNAN, P. J. Chemical synthesis of disaccharides of the specific phenolic glycolipid antigens from *Mycobacterium leprae* and of related sugars. Carbohydr. Res. 148 (1986) 287–297.
- FUJIWARA, T., HUNTER, S.-N., CHO, N.-S., ASPI-NALL, G. O. and BRENNAN, P. J. Chemical synthesis of disaccharides and trisaccharides of phenolic glycolipid antigens from the leprosy bacillus and preparation of a disaccharide protein conjugate for serodiagnosis of leprosy. Infect. Immun. 43 (1984) 245–252.
- FUJIWARA, T. and IZUMI, S. Synthesis of the neoglycoconjugates of phenolic glycolipid-related trisaccharides for the serodiagnosis of leprosy. Agric. Biol. Chem. 51 (1987) 1539–1547.
- FUJIWARA, T., IZUMI, S. and BRENNAN, P. J. Synthesis of 3,6-di-O-methylglucosyl disaccharides with methyl 3-(p-hydroyphenyl)propionate as a linker arm and their use in serodiagnosis of leprosy. Agric. Biol. Chem. 49 (1985) 2301–2308.
- GASTAMIBIDE-ODIER, M., LEDERER, E. and SARDA, P. Structure des aglycones des mycosides A et B. Tetrahedron Lett. 35 (1965) 3135–3143.
- GASTAMIBIDE-ODIER, M. and SARDA, P. Contribution à l'étude de la structure et de la biosynthèse de glycolipides spécifiques isolés de mycobactéries; les mycosides A et B. Pneumonologie 142 (1970) 241–255.

56, 1

Correspondence

115

- GERWIG, G. J., KAMERLING, J. P. and VLIE-GENTHART, J. F. G. Determination of the D and L configuration of neutral monosaccharides by highresolution capillary g.l.c. Carbohydr. Res. 62 (1978) 349–357.
- 14. GIGG, R., PAYNE, S. N. and CONANT, R. The allyl group for protection in carbohydrate chemistry. Part 14. Synthesis of 2,3-di-O-methyl-4-O-(3, 6di-O-methyl-β-D-glucopyranosyl)-L-rhamnopyranose (and its α-propyl glycoside): a haptenic por-

- tion of the major glycolipid from *Mycobacterium leprae*. J. Carbohydr. Chem. **2** (1983) 207–223.
- HUNTER, S. W. and BRENNAN, P. J. A novel pheolic glycolipid from *Mycobacterium leprae* possibly involved in immunogenicity and pathogenicity. J. Bacteriol. 147 (1981) 728–735.
- HUNTER, S. W., FUJIWARA, T. and BRENNAN, P. J. Structure and antigenicity of the major specific glycolipid antigen of *Mycobacterium leprae*. J. Biol. Chem. 257 (1982) 15072–15078.