

CORRESPONDENCE

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Infection by *M. leprae* is Governed by the Temperature at the Entry Point: a Preliminary Note

TO THE EDITOR:

The portal of entry of *Mycobacterium leprae*, the mode of transmission of leprosy in the community, and the evolution of the disease in an individual is still not clear. Previous studies conducted in nude mice have shown that *M. leprae* can be transmitted through unbroken nasal mucosa (^{1,3}) and abraded skin (²). It is well known that the nasal mucosa and feet are relatively cooler sites of the body. Infecting the nude mouse through intact and abraded skin at the flank was not successful (³). In this preliminary experiment an attempt is made to find out whether infection can take place if organisms are injected into the flanks of T900r mice which are almost similar to nude mice in their immunological status.

A group of 10 CBA mice were made immunodeficient using standard methodology except that the bone marrow transplantation was performed by injecting the harvested bone marrow into the peritoneal cavity (⁴). The *M. leprae* suspensions were obtained from harvesting the lepromatous nodules from the foot pads of a T900r mouse inoculated 15 months earlier with *M. leprae*. *M. leprae* suspensions were diluted so that 0.1 ml of a sample contained 10^7 *M. leprae*. In 10 T900r mice, hair was removed from both flanks and 0.1 ml of a suspension containing 10^7 bacilli was injected intradermally on both sides. Also, 0.03 ml of the bacillary suspension containing 10^4 bacilli

was injected into the hind foot pads of five normal CBA mice to confirm the viability of the bacilli. The animals were fed with standard food pellets and unlimited water. At 6, 12, and 15 months one animal from each group and two animals at 18 and 21 months were sacrificed. Three animals died due to unknown causes. The skin along with the subcutaneous tissue and muscle at the injected area on both sides were dissected out. The tissue from the left flanks was harvested and the acid-fast bacilli (AFB) counted. The tissue from the right flanks was immediately fixed in 10% buffered formalin and used for histopathological studies. The animals were autopsied and the organs were studied grossly and histopathologically. The tissues obtained at autopsy were also fixed in 10% buffered formalin and processed for 4- μ m-thick paraffin sections, and stained with hematoxylin and eosin (H&E) and a modified Fite-Faraco stain.

At 6, 9, 12, 18, and 21 months' growth up to 10^5 AFB was obtained from the foot pads of normal mice, confirming that the *M. leprae* used in this experiment were viable. The results of the bacterial count obtained from the T900r animals are given in The Table, showing a steady decrease in the number of organisms inoculated into the flanks.

Sections of skin from the flanks showed a diffuse granuloma occupying the dermis and the subcutaneous tissue including bun-

THE TABLE. *Bacterial counts from the flanks of T900r mice (one mouse was taken at months 6, 12, and 15 and two mice were taken at months 18 and 21).*

Months	Growth in skin of the flank (Group 1)
	Inoculum = 10^7
6	1×10^6
12	8.9×10^5
15	3.4×10^5
18	3.1×10^5
21	2.6×10^5
	5.3×10^4
21	3.9×10^4

dles of striated muscles. The granuloma consisted of macrophages, some with granular and some with vacuolated cytoplasm, lymphocytes, plasma cells and a few polymorphs (Fig. 1). They were mostly arranged in scattered clumps. Acid-fast staining showed macrophages filled with AFB. Almost all AFB were fragmented and granular (Fig. 2). AFB were not found in any other areas. The above changes were

seen consistently at 6 and 15 months. No histopathological changes were observed in the biopsy specimens obtained at month 12, 15 or 21. The hind foot, ears, liver, spleen, lungs, heart and kidneys obtained at autopsy from animals inoculated at the flanks showed no significant lesion except for a small, focal, nonspecific, mononuclear cell inflammatory infiltration of the liver and lungs. No acid-fast organisms were seen.

This study shows that when *M. leprae* were injected intradermally into the skin of the flanks of T900r mice they were taken up by the macrophages which aggregated in the injected sites. Clumps of macrophages containing AFB were seen at the injected site at 6 and 15 months. Morphologically, the AFB were all fragmented and granular. Their counts decreased from 10^7 to 10^6 at 6 months, 8.9×10^5 at 12 months, 3.4×10^5 at 15 months, 3.1×10^5 at 18 months and 3.9×10^4 at 21 months, clearly showing that there was no multiplication at the injected sites. There was also no evidence of dissemination of the infection to other parts of the body since autopsy of the animals did

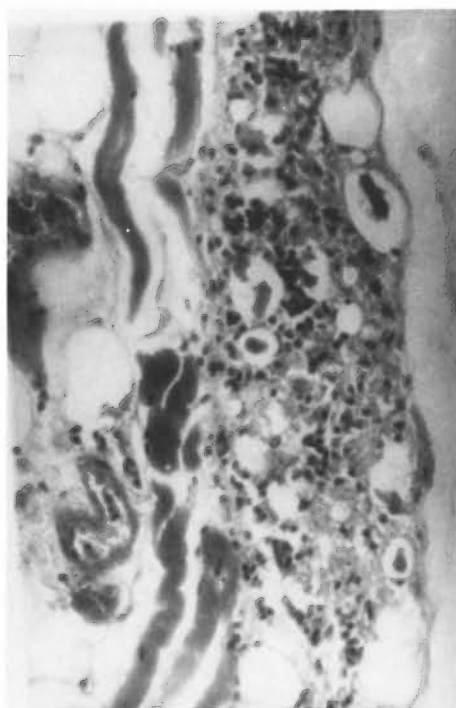


FIG. 1. Photomicrograph showing microgranuloma in the skin of the flank of T900r mice 6 months after inoculation with 10^7 viable *M. leprae* (H&E $\times 250$).

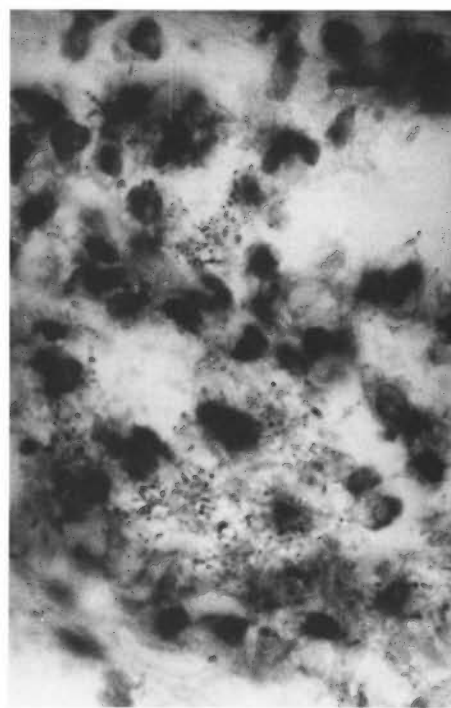


FIG. 2. Photomicrograph showing clumps of broken and granular AFB in macrophages of the granuloma shown in Figure 1 (modified Fite $\times 800$).

not show any organisms in organs such as the lungs, liver, spleen or ears.

One possible reason for the lack of multiplication may be that the skin at the flanks is warmer than that at the foot pads. It is reasonable, therefore, to suggest that although *M. leprae* may enter into the body through any skin site it can multiply and disseminate only in a suitable environment with a temperature less than normal core body temperature. The other reason may be that unlike the foot pad, the region of the flanks is a large area and the injected *M. leprae* may have fanned out without being localized and are thus lost to further evaluation.

The absence of granuloma or any histopathological changes at the right flanks of the animal examined at intervals of 12, 15, and 21 months, although AFB were harvested from the left side, is difficult to explain. The fact that even minimal inflammatory changes were absent suggests that the biopsy might have missed the site completely.

Further experiments are being done to confirm this study since the number of mice used here was small, and there were no control animals infected in the foot pads using the same inoculum and the same dosage for comparison.

—Gigi J. Ebenezer, M.D.

*Head, Department of
Histopathology and
Experimental Pathology*

—Shantha Arumugam

*Laboratory Technician
Department of Experimental Pathology*

—Charles K. Job, M.D., F.R.C.Path.,
F.A.M.S.

*Emeritus Scientist
Schieffelin Leprosy Research
and Training Center
Karigiri, Vellore District
Tamil Nadu, India 632 106*

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